FISEVIER

Contents lists available at ScienceDirect

European Journal of Pharmacology

journal homepage: www.elsevier.com/locate/ejphar



Pulmonary, Gastrointestinal and Urogenital Pharmacology

Characterization of the ileal muscarinic receptor system in 70-week-old type II Goto–Kakizaki diabetic rats; effects of cyclohexenonic long-chain fatty alcohol

Shin-ichi Okada ^a, Motoaki Saito ^{b,*}, Yukako Kinoshita ^b, Itaru Satoh ^b, Emi Kazuyama ^b, Atsushi Hayashi ^a, Keisuke Satoh ^b. Susumu Kanzaki ^a

- ^a Division of Pediatrics and Perinatology, Tottori University Faculty of Medicine, 36-1 Nishi-machi, Yonago, 683-8504, Japan
- b Division of Molecular Pharmacology, Tottori University Faculty of Medicine, 86 Nishi-machi, Yonago, 683-8503 Tottori, Japan

ARTICLE INFO

Article history:
Received 31 October 2008
Received in revised form 4 March 2009
Accepted 15 March 2009
Available online 24 March 2009

Keywords:
Type 2 diabetes
Muscarinic receptor
Cyclohexenonic long-chain fatty alcohol
Ileum
(Goto-Kakizaki (GK) rat)

ABSTRACT

As gastrointestinal motility disorders are frequently reported in patients with diabetes, we attempted to clarify the effects of cyclohexenonic long-chain fatty alcohol in type 2 Goto–Kakizaki (GK) diabetic enteropathy. At 40 weeks of age male GK rats divided into three groups (treated with 0, 2 or 8 mg/kg of cyclohexenonic long-chain fatty alcohol; started at the age of 40 weeks). Age-matched male Wistar rats were used in this study. At 70 weeks of age the ileal functions were estimated by organ bath studies using 100 mM KCl and carbachol. The expression levels of muscarinic M2 and M3 receptor mRNAs in the ileum were investigated by real-time polymerase chain reaction (PCR). Treatment with cyclohexenonic long-chain fatty alcohol did not alter the diabetic status of the GK rats, i.e., body weight, serum glucose, and serum insulin levels, but significantly ameliorated diabetes-induced hypercontractility of the rat ileum caused by carbachol in a dose-dependent manner. Although there were no significant differences in the expression levels of muscarinic M3 receptor mRNAs in any of the groups, cyclohexenonic long-chain fatty alcohol reversed the diabetes-induced up-regulation of intestinal muscarinic M2 receptor mRNAs in treatment groups. These results indicate that cyclohexenonic long-chain fatty alcohol exerts its therapeutic effects on hypercontractility in the ileum of 70-week-old GK type 2 diabetic rats by ameliorating overexpression of muscarinic M2 receptors.

© 2009 Elsevier B.V. All rights reserved.

1. Introduction

Gastrointestinal dysfunctions such as delayed gastric emptying, diarrhea, constipation, and abdominal pain occurs in many diabetic patients. These dysfunctions are primarily cause by diabetes-induced neuropathy (Hosking et al., 1978; Thomas and Eliasson, 1987), which upsets the balance between the cholinergic and adrenergic systems within the enteric nervous system, thereby disrupting the gastrointestinal tone and causing reduced peristalsis and dysmotility. The streptozotocin-induced diabetic rat, which is the most commonly used and well-investigated experimental model for type 1 diabetes, exhibits typical alterations of gastrointestinal function (Shinbori et al., 2006; Narimatsu et al., 2007). However, only limited information is available about enteropathy in rat models of type 2 diabetes.

The Goto–Kakizaki (GK) rat represents a spontaneous non-insulindependent diabetes model. GK rats are produced from normal Wistar rats by repetition of selective breeding, and are a widely accepted genetically determined rodent model for human type 2 diabetes. This genetic rat model is particularly relevant to human type 2 diabetes because it exhibits defects in glucose-stimulated insulin secretion, peripheral insulin resistance, and hyperinsulinemia as early as 4 weeks after birth, with impairment of insulin secretion and resulting hypoinsulinemia occurring at later stages (Goto et al., 1976; Goto and Kakizaki, 1981).

The tropical plant, Hygrophilia erecta Hochr., has been shown to contain some cyclohexenonic long-chain fatty alcohols that exert neurotrophic activities on cultured neurons from the cerebral cortex (Borg et al., 1997, 1990). Cyclohexenonic long-chain fatty alcohol has been found to directly increase the neurite extension as well as the biochemical differentiation of these neurons. In addition, we have reported that cyclohexenonic long-chain fatty alcohol prevented the progression of diabetes-induced alterations of the trachea, aorta, and bladder (Satoh et al., 2005; Shinbori et al., 2007; Saito et al., 2007). In other previous reports, we demonstrated that cyclohexenonic longchain fatty alcohol ameliorated the progression of diabetes-induced dysfunction of the ileum in streptozotocin-induced diabetic rats (Shinbori et al., 2006; Narimatsu et al., 2007). In the previous reports, although cyclohexenonic long-chain fatty alcohol did not improve the general features or serum glucose and insulin levels of the streptozotocin-induced diabetic rats, it significantly improved the thickness of the ileal wall and hypercontractility of the rat ileum in

^{*} Corresponding author. Department of Pathophysiological and Therapeutic Science, Division of Molecular Pharmacology, Tottori University Faculty of Medicine, 86 Nishimachi, Yonago 683-8503, Japan. Tel.: +81 859 38 6162; fax: +81 859 38 6160.

E-mail address: saitomo@grape.med.tottori-u.ac.jp (M. Saito).

Fig. 1. The chemical structure of cyclohexenonic long-chain fatty alcohol. 2.4.4trimethyl-3-(15-hydroxypentadecyl)-2-cyclohexen-1-one.

response to carbachol in a dose-dependent manner, and the improvement of hypercontractility was accomplished without qualitative alteration of the muscarinic receptor system (Shinbori et al., 2006; Narimatsu et al., 2007). From these reports, we propose that cyclohexenonic long-chain fatty alcohol may prevent not only type 1 diabetic peripheral neuropathy in the ileum, but also type 2 diabetes: furthermore, these effects appear to take place independently of increases of insulin and serum glucose.

Therefore, the aims of this study were to investigate type 2 diabetsinduced changes in the muscarinic receptor system in the rat ileum, and to clarify the effects of cyclohexenonic long-chain fatty alcohol on possible alterations of the muscarinic receptor system in the type 2 rat diabetic ileum.

2. Materials and methods

2.1. Animal models

All animal experiments were performed in accordance with the guidelines established by the Tottori University Committee for Animal Experimentation. Six-week-old male GK and Wistar rats were purchased from SLC (Shizuoka, Japan). All rats were kept under identical conditions, and had access to food and drinking water ad libitum. At the age of 12 weeks, seven Wistar and seven GK rats were used for preliminary functional studies. Then, the remaining rats were divided randomly into four groups (n = 6-8 each); a group of agematched Wistar rats treated with vehicle (A), a group of diabetic GK rats treated with vehicle (B), a group of diabetic GK rats treated with cyclohexenonic long-chain fatty alcohol, 2,4,4-trimethyl-3-(15-hydroxypentadecyl)-2-cyclohexen-1-one, at a daily dose of 2 mg/kg intramuscularly (C), and a group of diabetic GK rats treated with cyclohexenonic long-chain fatty alcohol at a daily dose of 8 mg/kg intramuscularly (D). Fig. 1 shows the chemical structure of this compound (Shinbori et al., 2007, 2008). A mixture of ethanol:saline: Tween 80 in the ratio 5:92.15:2.85 (the total volume was 1 ml/kg body weight) was used as a vehicle throughout this study (Saito et al., 2006, 2007; Shinbori et al., 2007, 2008). The drug or vehicle administrations were started at the age of 40 weeks in each animal. Upon reaching 70 weeks of the age, the rats were sacrificed with an overdose of pentobarbital (60 mg i.p.). Blood samples were collected from the vena cava, and the ileum was removed from each animal and placed in Krebs–Henseleit solution. These blood samples were frozen at $-80\,^{\circ}\text{C}$ until used. The rat ileum was immediately used for functional studies, and the rest of the tissue was frozen at $-80\,^{\circ}\text{C}$ for measurement of muscarinic M2 and M3 receptor mRNAs.

2.2. Serum glucose and insulin measurement

Serum glucose concentrations were measured by the hexokinase method using a Glucose C kit (Wako Pure Chemicals, Osaka, Japan) according to the manufacturer's instructions in all groups. The insulin concentrations were also measured by a Rat Insulin ELISA kit (Mercodia AB, Uppsala, Sweden) according to the manufacturer's instructions.

2.3. Measurement of contractile force in the isolated ileum

The functional studies were performed according to our previous reports (Shinbori et al., 2006; Narimatsu et al., 2007). The ileum was rinsed with Krebs-Henseleit solution, the most distal 10 cm discarded, and the remaining ileum cut into approximately 1 cm segments. The ileal segments were mounted longitudinally between a hook in the bottom of the muscle chamber and a force displacement transducer in an organ bath (25 ml) containing Krebs-Henseleit solution, and bubbled with 5% CO₂ and 95% O₂ (37 °C). One hook was suspended from a transducer (type 45196A; San-ei Instruments, Tokyo, Japan), and the lower hook was fixed to a plastic support leg to a micrometer (Mitutoyo, Tokyo, Japan). The ileum preparations were equilibrated unstretched for 30 min. A load of 1.0 g was applied to each strip by micrometer adjustment, and the load was readjusted to this level 30 min later. Changes in the tone of the strips were induced by means of a force transducer, and recorded on a personal computer (Macintosh G3; Apple Computer, Cupertino, CA) by use of Chart v 3.6.9 software and a PowerLab/16sp data acquisition system (AD Instruments, Castle Hill, Australia). Cumulative concentrationresponse curves to carbachol $(10^{-8} - 3 \times 10^{-5} \text{ M})$ and KCl (100 mM)were recorded.

2.4. Real-time polymerase chain reaction (PCR) of muscarinic M₂ and M₃ receptor mRNAs

Measurement of muscarinic receptor mRNAs was performed according to a method outlined in our previous reports (Shinbori et al., 2006; Narimatsu et al., 2007; Saito et al., 2007). Muscarinic M2 and M₃ receptor mRNAs in the experimental ileum were measured by real-time PCR. The mRNAs were purified using an RNeasy Mini Kit (Qiagen, Valencia, CA) according to the manufacturer's instructions. The reverse transcriptase (RT) mixture (20 ml) containing 2 mg of total RNA was prepared and incubated at 37 °C for 60 min according to a previously reported method (Shinbori et al., 2006; Narimatsu et al., 2007; Saito et al., 2007). Fifteen microliters of the mixture were used for real-time PCR, which was carried out using a Light Cycler thermal cycler system with a LightCycler-FastStart Hybridization Probe kit according to the manufacturer's instructions (Roche Diagnostics, Tokyo, Japan). The primers and probes were synthesized commercially by Nihon Gene Research Laboratories, Inc. (Sendai, Japan), and these sequences are shown in Table 1. The predicted product sizes of muscarinic M₂ and M₃ receptors were 148 bp and 149 bp, respectively. The primer and probe of the α -actin used were from the LightCycler-

Oligonucleotides of primers and hybridization probes for amplification of muscarinic

M ₂ and M ₃ receptors.					
Muscarinic M ₂ receptor subtype					
Primer	Forward 5' CCACTCCAGATGACAACT 3'				
	Reverse 5' GGCTACAACGTTCTGCTTT 3'				
Probe	5'-CCAACTAGTTCTACAGTGGTACTCGTTGGGGTG-3'-Fluorescein				
	3' Label: fluorescein				
	5'-LCRed640-ACACATCACCTTTTTGGGCCTTGGTGAC-3'-phosphorylation				
	5'Label: LCRed640 3' Label: phosphorylated				
Muscarini	c M₃ receptor subtype				

Forward 5' GGACTGTGGATGTGGAGAG 3' Primer Reverse 5' CGAGGAGTTGGTGTCAGA 3'

Probe 5'-CCAGAAGAGCATGGGTGATGGTGACAAC-3'-Fluorescein

3' Label: fluorescein

5'-LCRed 640-GTCAGAAGGATTTGACCAAGCTTCCCATCC-3'-phosphorylation5'Label: LCRed640 3' Label: phosphorylated

Primer/Probe Set (rat). α -actin was used as the internal standard. A total of 5 ml of solution was used for the sample.

2.5. Data analysis

The data for contractions induced by carbachol were normalized by those of 100 mM KCl. The EC₅₀ and $E_{\rm max}$ values were obtained by using a Macintosh computer (G3) loaded with Chart v3.6.9 software and a PowerLab/16sp data acquisition system. The expression of muscarinic M₂ and M₃ receptor mRNAs was quantified according to the expression of α -actin mRNAs in the experimental rat ileum. A statistical comparison of differences between groups was performed using analysis of variance and Fisher's multiple comparison tests. Values of P<0.05 were considered to indicate statistical significance.

2.6. Drugs and chemicals

Cyclohexenonic long-chain fatty alcohol, 2,4,4-trimethyl-3-(15-hydroxypentadecyl)-2-cyclohexen-1-one, was obtained from Meiji Milk Products Co., Ltd. (Tokyo, Japan). All other chemicals used were of reagent grade.

3. Results

3.1. General features of the experimental rats

The general features of the experimental rats are shown in Fig. 2 and Table 2. The GK diabetic rats showed significantly lower weight gain by the age of 10 weeks, as well as significantly lower weight gain during the experimental period compared with the controls (P<0.05) (Fig. 2). Significantly higher serum glucose levels were confirmed at the age of 12 weeks in the GK diabetic rats compared with the controls

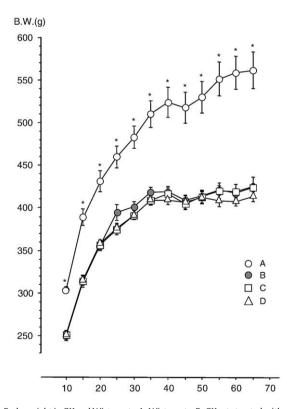


Fig. 2. Body weight in GK and Wistar rats. A: Wistar rats; B: GK rats treated with vehicle; C: GK rats treated with 2 mg/kg of cyclohexenonic long-chain fatty alcohol; and D: GK rats treated with 8 mg/kg of cyclohexenonic long-chain fatty alcohol. Data are shown as the means \pm S.E.M. of six to eight separate determinations in each group. *Significantly different from the other groups of age-matched rats.

Table 2General features of the experimental rats.

	Body weight (g	g)	_	
	40 weeks old	70 weeks old	Serum glucose (mg/dl)	Serum insulin (μg/L)
Α	525.7 ± 20.7	551.4 ± 22.3	132.2 ± 5.9	2.51 ± 0.33
В	419.3 ± 4.0^{a}	394.3 ± 3.7^{a}	208.1 ± 8.5^{a}	0.53 ± 0.10^{a}
C	416.4 ± 8.3^{a}	400.0 ± 8.7^{a}	217.3 ± 16.4^{a}	0.54 ± 0.10^{a}
D	409.3 ± 5.6^{a}	387.1 ± 8.1^{a}	209.1 ± 6.1^{a}	0.64 ± 0.15^{a}

A: control rats, B: diabetic rats, and C: diabetic rats treated with 2 mg/kg cyclohexenonic long-chain fatty alcohol. D: diabetic rats treated with 8 mg kg cyclohexenonic long-chain fatty alcohol. Data are shown as mean \pm S.E.M. of 6 to 8 separated determinations in each group.

 $(153.4\pm7.9\,$ and $233.1\pm12.3\,$ mg/dl for the Wistar and GK rats, respectively; P<0.05). The 70-week-old GK diabetic rats also showed significantly higher serum glucose and lower serum insulin levels than those of the control rats. Treatment with cyclohexenonic long-chain fatty alcohol did not improve the body weight gains during the observation periods, and had no effect on serum glucose or insulin levels.

3.2. Measurement of contractile force in the isolated ileum

The $E_{\rm max}$ and EC₅₀ values for the contractile responses of the ileal smooth muscles to carbachol (10^{-8} – 3×10^{-5} M) and KCl (100 mM) were determined (Table 3). At the age of 12 weeks, there was no significant difference in $E_{\rm max}$ values between Wistar and GK rats. However, significant hypercontractility to carbachol was observed in the ileum of 70-week-old GK rats. Diabetes-induced ileal hypercontractility to carbachol was significantly ameliorated by the high-dose (8 mg/kg) treatment with cyclohexenonic long-chain fatty alcohol. The EC₅₀ values obtained in response to carbachol indicated were not significantly difference among the four groups.

3.3. Real-time PCR analysis of muscarinic M₂ and M₃ receptor mRNAs

Fig. 3 shows the expressions of muscarinic M_2 and M_3 receptor mRNAs in the ileum as assessed by real-time PCR. In the untreated diabetes group (group B), the expressions of muscarinic M_2 receptor mRNAs were significantly higher than those in the controls (group A). Treatment with both doses of cyclohexenonic long-chain fatty alcohol significantly improved the up-regulation of muscarinic M_2 receptor mRNAs. The expressions of muscarinic M_3 receptor mRNAs in the untreated diabetes group, tended to be higher than those in agematched Wistar rats. However, because of the large variations in each group, these differences were not significantly different. Treatment with cyclohexenonic long-chain fatty alcohol tended to ameliorate diabetes-induced overexpression of muscarinic M_3 receptor mRNAs;

Table 3 Functional studies in the experimental rats.

	$E_{\rm max}/{ m KCl}$	$EC_{50}(10^{-7} \text{ M})$
12-week-old		
Wistar	1.36 ± 0.09	4.25 ± 0.81
GK	1.28 ± 0.06	4.37 ± 0.74
70-week-old		
A	1.28 ± 0.07	4.32 ± 0.87
В	1.60 ± 0.13^{a}	6.33 ± 1.46
C	1.52 ± 0.07^{a}	4.13 ± 0.43
D	1.36 ± 0.05	4.65 ± 1.18

The $E_{\rm max}$ values to the contractile response to carbahcol and KC1 (100 mM) were determined. The EC₅₀ values with respect to carbachol. A: control rats, B: diabetic rats, C: diabetic rats treated with 2 mg/kg cyclohexenonic long-chain fatty alcohol. D: diabetic rats treated with 8 mg/kg cyclohexenonic long-chain fatty alcohol. Data are shown as mean \pm S.E.M. of 6 to 8 separated determinations in each group.

^a Significantly different from group A. (P<0.05).

^a Significantly different from group A (P<0.05).

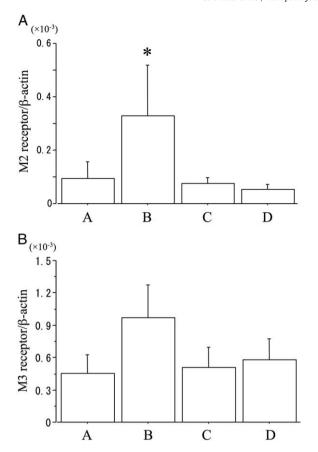


Fig. 3. A and B. Expressions of muscarinic M_2 and M_3 receptor mRNAs, respectively. A: Wistar rats; B: GK rats treated with vehicle; C: GK rats treated with 2 mg/kg of cyclohexenonic long-chain fatty alcohol; and D: GK rats treated with 8 mg/kg of cyclohexenonic long-chain fatty alcohol. All data were normalized by b-actin. Data are shown as the means \pm S.E.M. of six to eight separate determinations in each group. *Significantly different from the other groups of age-matched rats.

the expression levels of muscarinic M₃ receptor mRNAs in GK rats treated with both doses were close to that of control Wistar rats. However, no significant differences in the expressions of muscarinic M₃ receptor mRNAs were found among the four groups.

4. Discussion

Several studies have examined contractile responses of ileal smooth muscle to acetylcholine or cholinergic agonists in streptozotocin-induced diabetes, and have alternatively reported that the responses are elevated (Shinbori et al., 2006; Narimatsu et al., 2007; Carrier and Aronstam, 1990; Anjaneyulu and Ramarao, 2002; Talubmook et al., 2003) or decreased (Lucas and Sardar, 1991; Coulson et al., 2002). Interestingly, Daneshgari et al. (2006) reported that diabetic bladders may undergo a transition from a compensated to a decompensated state and that this transition in the streptozotocin rat model may begin at 9 to 12 weeks after the induction of diabetes. It may be that the same phenomenon occurs in the streptozotocin-induced diabetic rat ileum, and thus the conflicting results described above may have been due to the durations of the diabetic periods.

However, the majority of human diabetes is type 2. There are some reports indicating that neuropathy in type 1 diabetes is different from that in type 2 diabetes (Schmidt et al., 2003, 2004; Kamiya et al., 2005), and only limited information is available in regard to ileal dysfunction in type 2 diabetic models. In the present study, although there was no significant difference of $E_{\rm max}$ values for carbachol between Wistar and GK rats at the age of 12 weeks, we clearly demonstrated the hypercontractility of detrusor smooth muscle in response to carbachol

in GK rats at the age of 70 weeks compared to age-matched Wistar rats. These data were similar to those for streptozotocin-induced diabetic rats previously reported (Shinbori et al., 2006; Narimatsu et al., 2007). These data are particularly interesting because early-stage streptozotocin-induced diabetic rats and late-stage GK rats show similar patterns of ileal dysfunction. In previous studies, we showed that significant alterations of the ileal muscarinic receptor system occurred over the eight-week period of streptozotocin-induced diabetes (Shinbori et al., 2006; Narimatsu et al., 2007). Thickening the ileal wall, hypercontractility in response to carbachol, and up-regulations of the expressions of muscarinic M_2 and M_3 receptor mRNAs were observed in the streptozotocin-induced diabetic rat ileum. These alterations of the muscarinic receptor system in the rat ileum were quantitative rather than qualitative pharmacologically and biologically (Shinbori et al., 2006; Narimatsu et al., 2007).

As regards the role of muscarinic receptors, we have suggested that the rat ileum undergoes contractions mainly via the muscarinic M₃ receptor subtype using pharmacological methods (Narimatsu et al., 2007). However, based on the results of previous studies, we cannot rule out the possibility that the muscarinic M₂ receptor subtype does not directly contribute to cholinergic contractions. In addition, Unno et al. (2006) reported results for muscarinic M2-knockout, muscarinic M₃-knockout, muscarinic M₂/M₃-double knockout, and wild-type mice, demonstrating that muscarinic M₂ and M₃ receptors participate in mediating cholinergic contractions in the mouse ileum, with the latter subtype of receptors assuming a substantial role in this process. In order to clarify the mechanisms underlying type 2 diabetic enteropathy, we undertook to measure the expression levels of muscarinic M2 and M3 receptor mRNAs utilizing real-time PCR methods. In this study, we found that muscarinic M2 receptor mRNAs were significantly up-regulated, and muscarinic M₃ receptor mRNAs were slightly up-regulated in the ileum of 70-week-old GK type 2 diabetic rats. The lack of statistical significance in the latter result may have been due to the small size of the groups we studied. The possible mechanisms of this up-regulation of muscarinic receptors are thought to include a decrease in cholinergic nerve density, or a defect in the neurotransmitter release mechanism (Miyamae et al., 2004; Tong et al., 1996). The diabetes-associated neuropathy may inhibit the release of acetylcholine from cholinergic nerves, in turn inducing the overexpression of muscarinic receptors in the diabetic ileal smooth muscle. Such overexpression may enhance signaling downstream of these receptors and may increase smooth muscle contraction, according to the results of the present organ bath study.

Cyclohexenonic long-chain fatty alcohol was first reported to have neurotrophic activities on cultured neurons from the cerebral cortex (Borg et al., 1997, 1990). The C26-alcohol, N-hexacosanol, has been found to directly increase neurite extension as well as biochemical differentiation of these neurons. Watanabe and Miyagawa reported that cyclohexenonic long-chain fatty alcohol has beneficial effects on peripheral neuropathy and cystopathy in streptozotocin-induced diabetic rats (Watanabe and Miyagawa, 2002). They also demonstrated that diabetes-induced alteration of motor sciatic nerve conduction is normalized by treatment with cyclohexenonic longchain fatty alcohol (Watanabe and Miyagawa, 2002). In more recent studies, our group has investigated mainly therapeutic and/or preventive effects on streptozotocin-induced cystopathy, enteropathy, angiopathy, nephropathy, and tracheal dysfunction (Narimatsu et al., 2007; Okada et al., 2008; Saito et al., 2006, 2007; Shinbori et al., 2006, 2007; Suzuki et al., 2006). We found that cyclohexenonic long-chain fatty alcohol has no effect on serum insulin or glucose levels in streptozotocin-induced diabetic animals, but that it has therapeutic and/or preventive effects on streptozotocin-induced cystopathy, enteropathy, angiopathy, nephropathy, and tracheal dysfunction. Recently, we have also reported that streptozotocin-induced diabetes caused alterations of amino acid components in the central nervous system of rats, and that treatment with cyclohexenonic long-chain

fatty alcohol significantly ameliorated these alterations (Shinbori et al., 2008). These reports strongly suggest that cyclohexenonic long-chain fatty alcohol has therapeutic and/or preventive effects on not only peripheral neuropathy but also central neuropathy induced by streptozotocin. In this study, we also investigated the therapeutic effects of cyclohexenonic long-chain fatty alcohol on type 2 diabetesinduced ileal alterations. We clearly demonstrated that cyclohexenonic long-chain fatty alcohol significantly ameliorated diabetes-induced hypercontractility of the rat ileum in response to carbachol, and that cyclohexenonic long-chain fatty alcohol reversed the diabetes-induced up-regulation of intestinal muscarinic M₂ receptor mRNAs in the rats. From this study, although the precise mechanisms are unclear, it is clear that cyclohexenonic long-chain fatty alcohol reduces hypercontractility of the ileum of 70-week-old GK type 2 diabetic rats by ameliorating overexpression of muscarinic M2 receptor proteins and mRNAs. The physiological and pharmacological role of cyclohexenonic long-chain fatty alcohol in type 2 diabetes mellitus, however, remains unclear and warrants further study.

In conclusion, our data indicate that cyclohexenonic long-chain fatty alcohol ameliorates hypercontractility in the ileum of 70-week-old GK type 2 diabetic rats by ameliorating overexpression of muscarinic M₂ receptor and possibly M₃ receptor subtypes.

References

- Anjaneyulu, M., Ramarao, P., 2002. Studies on gastrointestinal tract functional changes in diabetic animals. Methods Find Exp. Clin. Pharmacol. 24, 71–75.
- Borg, J., Kesslak, P.J., Cotman, C.W., 1990. Peripheral administration of a long-chain fatty alcohol promotes septal cholinergic neurons survival after fimbria–fornix transection. Brain Res. 518, 295–298.
- Borg, J., Toazara, J., Hietter, H., Henry, M., Schmitt, G., Luu, B., 1997. Neurotrophic effect of naturally occurring long-chain fatty alcohols on cultured CNS neurons. FEBS Lett. 213, 406–410.
- Carrier, G.O., Aronstam, R.S., 1990. Increased muscarinic responsiveness and decreased muscarinic receptor content in ileal smooth muscle in diabetes. J. Pharmacol. Exp. Ther. 254, 445–449.
- Coulson, F.R., Jacoby, D.B., Fryer, A.D., 2002. Increased function of inhibitory neuronal M2 muscarinic receptors in trachea and ileum of diabetic rats. Br. J. Pharmacol. 135, 1355–1362
- Daneshgari, F., Liu, G., Imrey, P.B., 2006. Time dependent changes in diabetic cystopathy in rats include compensated and decompensated bladder function. J. Urol. 176, 380–386.
- Goto, Y., Kakizaki, M., 1981. The spontaneous-diabetes rat: a model of noninsulin dependent diabetes mellitus. Proc. Jpn. Acad. 57, 381–384.
- Goto, Y., Kakizaki, M., Masaki, N., 1976. Production of spontaneous diabetic rats by repetition of selective breeding. Tohoku J. Exp. Med. 119, 85–90.
- Hosking, D.J., Bennett, T., Hampton, J.R., 1978. Diabetic autonomic neuropathy. Diabetes 27, 1043–1055.
- Kamiya, H., Murakawa, Y., Zhang, W., Sima, A.A., 2005. Unmyelinated fiber sensory neuropathy differs in type 1 and type 2 diabetes. Diabetes Metab. Res. Rev. 21, 448–458.

- Lucas, P.D., Sardar, A.M., 1991. Effects of diabetes on cholinergic transmission in two rat gut preparations. Gastroenterology 100, 123–128.
- Miyamae, K., Yoshida, M., Inadome, A., Murakami, S., Otani, M., Iwashita, H., Masunaga, K., Ueda, S., 2004. Acetylcholine release from urinary bladder smooth muscles of non-insulin-dependent diabetic rats. Urol. Int. 73, 74–80.
- Narimatsu, N., Saito, M., Kazuyama, E., Hisadome, Y., Kinoshita, Y., Satoh, I., Okada, S., Suzuki, H., Yamada, M., Satoh, K., 2007. N-hexacosanol prevents diabetes-induced rat ileal dysfunction without qualitative alteration of the muscarinic receptor system. Biomed. Res. 28, 267–273.
- Okada, S., Saito, M., Kazuyama, E., Hanada, T., Kawaba, Y., Hayashi, A., Satoh, K., Kanzaki, S., 2008. Effects of N-hexacosanol on nitric oxide synthase system in diabetic rat nephropathy. Mol. Cell. Biochem. 315, 169–177.
- Saito, M., Kinoshita, Y., Satoh, I., Shinbori, C., Kono, T., Hanada, T., Uemasu, J., Suzuki, H., Yamada, M., Satoh, K., 2006. N-hexacosanol ameliorates streptozotocin-induced diabetic rat nephropathy. Eur. J. Pharmacol. 544, 132–137.
- Saito, M., Kinoshita, Y., Satoh, I., Shinbori, C., Suzuki, H., Yamada, M., Watanabe, T., Satoh, K., 2007. Ability of cyclohexenonic long-chain Fatty alcohol to reverse diabetes-induced cystopathy in the rat. Eur. Urol. 51, 479–488.
- Satoh, I., Saito, M., Kinoshita, Y., Shomori, K., Suzuki, H., Yamada, M., Kono, T., Satoh, K., 2005. Effects of cyclohexenonic long-chain fatty alcohol on diabetic rat trachea. Life Sci. 77, 2030–2039.
- Schmidt, R.E., Dorsey, D.A., Beaudet, L.N., Peterson, R.G., 2003. Analysis of the Zucker Diabetic Fatty (ZDF) type 2 diabetic rat model suggests a neurotrophic role for insulin/IGF-I in diabetic autonomic neuropathy. Am. J. Pathol. 163, 21–28.
- Schmidt, R.E., Dorsey, D.A., Beaudet, L.N., Parvin, C.A., Zhang, W., Sima, A.A., 2004. Experimental rat models of types 1 and 2 diabetes differ in sympathetic neuroaxonal dystrophy. J. Neuropathol. Exp. Neurol. 63, 450–460.
- Shinbori, C., Saito, M., Kinoshita, Y., Satoh, I., Kono, T., Nanba, E., Adachi, K., Suzuki, H., Yamada, M., Satoh, K., 2006. N-hexacosanol reverses diabetic induced muscarinic hypercontractility of ileum in the rat. Eur. J. Pharmacol. 545, 177–184.
- Shinbori, C., Saito, M., Kinoshita, Y., Satoh, I., Kono, T., Hanada, T., Nanba, E., Adachi, K., Suzuki, H., Yamada, M., Satoh, K., 2007. Cyclohexenonic long-chain fatty alcohol has therapeutic effects on diabetes-induced angiopathy in the rat aorta. Eur. J. Pharmacol. 567, 139–144.
- Shinbori, C., Shirayama, Y., Mitani, H., Saito, M., Satoh, K., 2008. Effects of cyclohexanonic long-chain fatty alcohol, tCFA15 on amino acids in diabetic rat brain: a preliminary study. Neurochem. Res. 33, 1189–1195.
- Suzuki, H., Saito, M., Kinoshita, Y., Satoh, I., Kono, T., ShinBori, C., Anastasios, S., Yamada, M., Satoh, K., 2006. Preventive effects of cyclohexenonic long-chain fatty alcohol on diabetic cystopathy in the rat. Can. J. Physiol. Pharm. 84, 195–201.
- Talubmook, C., Forrest, A., Parsons, M., 2003. Streptozotocin-induced diabetes modulates presynaptic and postsynaptic function in the rat ileum. Eur. J. Pharmacol. 469, 153–158.
- Thomas, P.K., Eliasson, S.G., 1987. Diabetic neuropathy. In: Duck, P.J., Thomas, P.K., Lambert, E.H. (Eds.), Peripheral Neuropathy. WB. Saunders, Philadelphia, PA, pp. 1777–1810.
- Tong, Y.C., Hung, Y.C., Lin, S.N., Cheng, J.T., 1996. Alterations in urinary bladder synaptosomal neurotransmitter concentrations in two-week streptozotocininduced diabetic rats. Neurosci. Lett. 206, 165–168.
- Unno, T., Matsuyama, H., Izumi, Y., Yamada, M., Wess, J., Komori, S., 2006. Roles of M₂ and M₃ muscarinic receptors in cholinergic nerve-induced contractions in mouse ileum studied with receptor knockout mice. Br. J. Pharmacol. 149, 1022–1030.
- Watanabe, T., Miyagawa, I., 2002. Effects of long-chain fatty alcohol on peripheral nerve conduction and bladder function in diabetic rats. Life Sci. 70, 2215–2224.